



*Int. J. Modn. Res. Revs.*

Volume 3, Issue 8, pp 739-743, August, 2015

ISSN: 2347-8314

**ORIGINAL ARTICLE**

**IMPACT OF *Pimenta dioica* LEAF EXTRACT ON CERTAIN BLOOD PARAMETERS IN STZ INDUCED DIABETIC WISTAR RATS**

**K. Yogalakshmi and \*J. Vaidehi**

Department of Zoology, Faculty of Science, Annamalai University,  
Annamalai nager - 608 002, Tamil nadu, India

*Article History: Received 20<sup>th</sup> July, 2015, Accepted 16<sup>th</sup> August, 2015, Published 17<sup>th</sup> August, 2015*

**ABSTRACT**

The present study was designed to investigate the certain blood parameters in STZ induced diabetic rats. The methanolic leaf extract of *Pimenta dioica* at the dose of 75 and 150 mg/kg of body weight was administered orally once in a day to the diabetic induced group for 45 days. Glibenclamide (0.6 mg/kg of body weight) was used as reference drug. In the present study body weight was significantly ( $P < 0.05$ ) decreased in diabetic rats when compared with that of the normal control rats. In diabetic rats there was a significant increase in the level of plasma glucose and significant decrease in the plasma insulin. Oral administration of *Pimenta dioica* (75 mg and 150 mg/kg.bw) and glibenclamide (0.6 mg/kg. bw) to diabetic rats significantly ( $P < 0.05$ ) increased the body weight and plasma insulin and markedly decreased the plasma glucose level when compared with that of the diabetic control rats. The diabetic rats showed a significant decrease in the level of hemoglobin and significant ( $P < 0.05$ ) increase in the level of HbA<sub>1c</sub>. Whereas the levels of plasma urea, plasma uric acid and creatinine significantly increased in the diabetic control group when compared with that of the normal control group. The level of hemoglobin after the administration of *Pimenta dioica* (75mg and 150 mg/kg.bw) and glibenclamide (0.6mg/kg.bw) were significantly increased in the diabetic rats. The level of plasma urea, uric acid and creatinine after orally administering *Pimenta dioica* (75mg and 150 mg/kg.bw) significantly decreased in the diabetic control group. Thus the present study suggested significant blood parameters potential in the methanolic leaf extracts of *P. dioica*.

**Keywords:** *Pimenta dioica*, Glibenclamide, blood parameters, methanolic leaf extract.

**1. INTRODUCTION**

Diabetes mellitus is a metabolic disease characterized by high blood glucose level resulting from defects in insulin secretion, insulin action or both [Khan et al., 2009]. It is a chronic disorder that affects the metabolism of carbohydrates, fats, proteins and electrolytes in the body, leading to severe complications which are classified into acute, sub-acute and chronic [Rang et al., 1991]. Acute complications include hypoglycemia, diabetic ketoacidosis, hyperglycemic non-ketotic syndrome [Kmentz and Natras, 1991] while sub-acute complications include thirst, polyuria, lack of energy, visual blurriness and weight loss [Kumar and Clark, 2002]. The management of diabetes is considered a global problem and a cure has yet to be discovered despite many great strides have been made in understanding the management of diabetes. Despite the presence of known antidiabetic medicine in the pharmaceutical market, diabetes and the related complications continued to be a major medical problem. The

searching for new antidiabetic drugs from natural plants is still attractive because they contain substances which take alternative and safe effect on diabetes mellitus. Concurrently, Phytochemicals identified from traditional medicinal plants are presenting exciting opportunities for the development of new drug therapies. Currently available drug regimes for the management of diabetes mellitus have certain drawbacks. Recently there has been increasing interest in the use of medicinal plants [Pulok and Mukherjee 2002; Alluri and Krishnaraj, 2006]. In many countries traditional plants are used to control diabetes. Plants play a major role in the discovery of new therapeutic agents and have received much attention as sources of biologically active substances including antioxidants, hypoglycemic and hypolipidemic agents [Marles et al., 1995]. Many useful plants and herbs introduced in pharmacological and clinical trials have confirmed their blood sugar lowering effect. So it is essential to know about the pharmacological evaluation of various plants used in the traditional system of medicine [Gupta et al., 2007]. Spices and herbs have been added to foods since ancient times, not only as flavouring agents, but also as folk medicines and food preservatives [Beucha, 1994; Nakatani, 1994; Cutler, 1995]. *Pimenta dioica* is one

\*Corresponding author: **Dr. J. Vaidehi**, Department of Zoology,  
Faculty of Science, Annamalai University,  
Annamalai nager - 608 002, Tamil nadu, India

such plant used as a flavouring agent which was also later diagnosed for its antioxidant, antidiabetic, anti-inflammatory and anti-microbial properties. The whole plant exhibit medicinal values. The dried leaves contain 0.7 to 2.9 % of oil which is called pimento oil. Like berry oil it contains eugenol as its main constituent but has an inferior odour and flavour. The existences of antioxidants are beneficial in preventing disease complexes such as cardiovascular, diabetes, cancer, rheumatoid arthritis, inflammatory bowel pancreatitis, hematological and neurodegenerative diseases [Irshad and Chaudhuri, 2002; Craig, 1997]. In this context, the present study was aimed to investigate the antidiabetic activity of the methanolic leaf extract of *Pimenta dioica* in streptozotocin induced diabetic rats.

## 2. MATERIALS AND METHODS

### Chemicals

Streptozotocin (STZ) was purchased from Sigma-Chemical Co. Bangalore. All other chemicals and reagents used for this study were of analytical grade.

### Plant material

*Pimenta dioica* was collected from Kumuli, Kerala State, India.

### Preparation of extract

The *Pimenta dioica* leaves were dried at room temperature and then were powdered using dry grinder and passed through sieve. Hundred grams of *Pimenta dioica* leaves were packed in a Soxhlet apparatus and extracted with methanol. The methanol extracts were concentrated on a rotary evaporator.

### Experimental animals

Male Wistar albino rats (150-200 g) were procured from Rajah Muthiah Medical College and Hospital, Annamalai University, Chidambaram, Tamilnadu, India and were housed in polycarbonate cages in an animal room with 12 hours day – night cycle. The animals were allowed free access to tap water and standard laboratory rat food. The animal treatment and protocol employed were approved by the TAEC, Annamalai University, Annamalai Nagar, India (Registration Number - 1084/2014/CPCSEA)

### Induction of experimental diabetes

Diabetes was induced in rats by intraperitoneal (I.P.) injection of streptozotocin (STZ) at a dose of 55 mg/kg b.w dissolved in 0.1 M cold citrate buffer (pH = 4.5) [Bandaranayake, 2002]. The rats were allowed to drink 5% glucose solution overnight to overcome the drug-induced hypoglycemia. The blood glucose values above 250 mg/dl on the third day after streptozotocin injection were considered as diabetic rats. Then the treatment was started on the fifth day after streptozotocin injection and it was considered as the first day of treatment.

### Experimental design

All animals were randomly divided into five groups with six animals in each group

1. Normal untreated rats
2. Diabetic control rats (STZ 55 mg/kg of body weight).
3. Diabetic rats treated with methanolic extract of *Pimenta dioica* leaves (75 mg/kg of body weight)
4. Diabetic rats treated with methanolic extract of *Pimenta dioica* leaves (150 mg/kg of body weight)
5. Diabetic rats treated with standard drug, glibenclamide (0.6 µg/kg of body weight).

### Analytical procedures

The estimation of blood glucose was carried out by the method O-toluidine using the modified reagent of [Sasaki et al., 1972]. The estimation of hemoglobin was done by the method of [Drabkin and Austin, 1932]. The glycosylated hemoglobin in the blood was estimated by the protocol of [Sudhakar Nayak and Pattabiraman 1981]. The plasma insulin was assayed by ELISA method (Enzyme Linked Immunosorbant Assay) using Boehringer Mannheim Kit (Boehringer analyzer, ES 300). Urea in the plasma was estimated by using the diagnostic kit based on the method of [Fawcett and Scott, 1960]. Uric acid was estimated by the method adapted by Caraway [1955]. Creatinine in the plasma was estimated by using the diagnostic kit based on the protocol of Tietz [1987] using [Jaffe, 1886] colour reaction.

### Statistical Analysis

All data are expressed as mean ± S.E. Statistical analysis was performed using one-way ANOVA followed by Tukey's multiple tests using SPSS (version 18) computer software. In all cases, P-value of less than 0.05 was considered to be significant.

## 3. RESULTS

In the present study, the changes in the body weight, plasma glucose and plasma insulin in control and experimental animals were presented in Table-1. Body weight was significantly ( $P < 0.05$ ) decreased in diabetic rats when compared with that of the normal control rats. In diabetic rats there was a significant increase in the level of plasma glucose and significant decrease in the plasma insulin. Oral administration of *Pimenta dioica* (75 mg and 150 mg/kg.bw) and glibenclamide to diabetic rats significantly ( $P < 0.05$ ) increased the body weight and plasma insulin and markedly decreased the plasma glucose level when compared with that of the diabetic control rats.

The level of hemoglobin, glycosylated hemoglobin (HbA<sub>1c</sub>), urea, uric acid and creatinine in normal control and experimental animals are shown in Table – 2. The diabetic rats showed a significant decrease in the level of hemoglobin and significant ( $P < 0.05$ ) increase in the level of HbA<sub>1c</sub>. Whereas the levels of plasma urea, plasma uric acid and creatinine significantly increased in the diabetic control

**Table 1: Effect of *Pimenta dioica* on body weight, plasma glucose and plasma insulin in normal control and diabetic rats.**

| Groups                             | Body weight (g)                |                                | Plasma glucose (mg/dl)          | Plasma Insulin ( $\mu$ U/ml)  |
|------------------------------------|--------------------------------|--------------------------------|---------------------------------|-------------------------------|
|                                    | Initial (0 day)                | Final (45 days)                |                                 |                               |
| Control                            | 182.89 $\pm$ 4.10 <sup>c</sup> | 210.03 $\pm$ 3.81 <sup>a</sup> | 86.71 $\pm$ 4.50 <sup>d</sup>   | 15.80 $\pm$ 1.08 <sup>a</sup> |
| Diabetic                           | 184.0 $\pm$ 4.20 <sup>b</sup>  | 140.45 $\pm$ 4.66 <sup>c</sup> | 287.78 $\pm$ 11.45 <sup>a</sup> | 7.40 $\pm$ 0.31 <sup>e</sup>  |
| Diabetic + <i>P. dioica</i> (M)/75 | 178.51 $\pm$ 3.30 <sup>d</sup> | 190.09 $\pm$ 4.40 <sup>c</sup> | 135.24 $\pm$ 5.84 <sup>b</sup>  | 14.21 $\pm$ 0.70 <sup>d</sup> |
| Diabetic+ <i>P. dioica</i> (M)/150 | 172.25 $\pm$ 4.21 <sup>e</sup> | 187.51 $\pm$ 2.30 <sup>d</sup> | 105.23 $\pm$ 8.81 <sup>c</sup>  | 15.31 $\pm$ 1.20 <sup>b</sup> |
| Glibenclamide (0.6 $\mu$ g)        | 185.48 $\pm$ 4.50 <sup>a</sup> | 206.00 $\pm$ 3.80 <sup>b</sup> | 97.54 $\pm$ 6.80 <sup>e</sup>   | 15.08 $\pm$ 1.15 <sup>c</sup> |

Values are given as mean  $\pm$  SD (n=6 rats) significantly different at P<0.05 when compared with control group

**Table 2: Effect of *Pimenta dioica* on haemoglobin, glycosylated haemoglobin, urea, uric acid and creatinine in normal control and diabetic rats**

| Groups                             | Hemoglobin (g/dl)             | Glycosylated hemoglobin (HbA <sub>1c</sub> ) % | Urea (mg/dl)                  | Uric acid (mg/dl)            | Creatinine (mg/dl)            |
|------------------------------------|-------------------------------|--|-------------------------------|------------------------------|-------------------------------|
| Control                            | 13.06 $\pm$ 1.02 <sup>a</sup> | 0.42 $\pm$ 0.01 <sup>e</sup>                   | 24.90 $\pm$ 1.80 <sup>e</sup> | 1.29 $\pm$ 0.08 <sup>e</sup> | 0.43 $\pm$ 0.030 <sup>e</sup> |
| Diabetic                           | 9.08 $\pm$ 0.46 <sup>e</sup>  | 1.38 $\pm$ 0.10 <sup>a</sup>                   | 44.41 $\pm$ 3.40 <sup>a</sup> | 2.29 $\pm$ 0.19 <sup>a</sup> | 0.82 $\pm$ 0.04 <sup>a</sup>  |
| Diabetic + <i>P. dioica</i> (M)/75 | 11.01 $\pm$ 0.84 <sup>d</sup> | 0.49 $\pm$ 0.03 <sup>b</sup>                   | 31.71 $\pm$ 2.91 <sup>b</sup> | 1.79 $\pm$ 0.13 <sup>b</sup> | 0.52 $\pm$ 0.08 <sup>b</sup>  |
| Diabetic+ <i>P. dioica</i> (M)/150 | 12.04 $\pm$ 1.06 <sup>c</sup> | 0.47 $\pm$ 0.05 <sup>c</sup>                   | 30.33 $\pm$ 2.35 <sup>c</sup> | 1.71 $\pm$ 0.14 <sup>c</sup> | 0.49 $\pm$ 0.11 <sup>d</sup>  |
| Glibenclamide (0.6 $\mu$ g)        | 12.75 $\pm$ 1.13 <sup>b</sup> | 0.45 $\pm$ 0.03 <sup>d</sup>                   | 29.17 $\pm$ 2.21 <sup>d</sup> | 1.64 $\pm$ 0.01 <sup>d</sup> | 0.45 $\pm$ 0.12 <sup>c</sup>  |

Values are given as mean  $\pm$  SD (n=6 rats), significantly different at P<0.05 when compared with control group

group when compared with that of the normal control group. The level of hemoglobin after the administration of *Pimenta dioica* (75mg and 150 mg/kg.bw) and glibenclamide were significantly increased in the diabetic rats. The level of plasma urea, uric acid and creatinine after orally administering *Pimenta dioica* (75mg and 150 mg/kg.bw) significantly decreased in the diabetic control group.

#### 4.DISCUSSION

The loss of body weights observed in STZ induced diabetic rat group (after a period of 30 days) may be due to muscle wasting and loss of tissue proteins upon the induction of diabetes with STZ [Wanston et al., 1990; Chatterjee and Shinde,2002]. Earlier [Santhakumar et al., 2006; Kaleem et al., 2005] reported 26% and 52% reduction in the FBG levels of diabetic rats treated with aqueous extracts of *Piper betel* leaves and *Piper nigrum* seeds respectively for 30 days. Glycosylated hemoglobin is used as a marker for estimating the degree of protein glycation in diabetes mellitus. HbA<sub>1c</sub> was found to increase in patients with diabetes mellitus and the amount of increase is directly proportional to the fasting blood glucose level [Al-yassin and Ibrahim,1981]. In diabetic condition, the excess glucose present in the blood reacts with hemoglobin to form HbA<sub>1c</sub> [Koenig et al., 1976]. Estimation of HbA<sub>1c</sub> has been known to be particularly useful in monitoring the effectiveness of therapy in diabetes. The observed increase in the levels of HbA<sub>1c</sub> in diabetic control group rats was due to the presence of excessive amounts of blood glucose. During diabetes as the excess of glucose present in blood reacts with Hb to form HbA<sub>1c</sub> the total HB level was observed to be decreased in

diabetic rats [Saudek et al., 2006]. The decrease in body weight with diabetes mellitus has been attributed to the gluconeogenesis i.e., catabolism of proteins and fats, which is associated with the characteristic loss of body weight due to increased muscle wasting and loss of tissue proteins [Shirwaikar et al., 2004; Shirwaikar et al., 2006] However, in diabetic state, lipoprotein lipase is not activated due to insulin resistance deficiency, resulting in hypertriglyceridemia and insulin deficiency is also associated with hypercholesterolemia due to metabolic abnormalities [Jarald et al., 2008]. Administration of *Acorus calamus* methanolic extract to diabetic rats showed a significant decrease in the fasting blood glucose level and an increase in the serum insulin levels and may be due to the presence of saponins, glycosides and sequiterpenoids which possesses hypoglycemic property [Parab and Mengi,2002; Campos et al., 2009; Gengaihi et al., 2011].

The diabetic hyperglycemia induces elevation of the plasma levels of urea, uric acid and creatinine which are considered as significant markers in renal function [Almdal ,1988]. Catabolism of the protein and nucleic acids results in the formation of urea. In diabetic condition the amino acids breakdown result in an increased production of urea [Chattopadhyay and Bandyopadhyay,2005]. Diabetes mellitus impairs the normal capacity of the liver to synthesize glycogen. The regulation of glycogen metabolism in vivo occurs by the enzymes glycogen synthase and glycogen phosphorylase. The reduced glycogen store in the diabetic rats has been attributed to the reduced activity of glycogen synthase and increased activity of glycogen phosphorylase. This is probably due to the lack of insulin in the diabetic state, which results in the inactivation of the

glycogen synthetase systems [Shirwaikar, 2006]. Glucose is transported out of the liver to increase the blood glucose concentration. Normally insulin inhibits the hepatic glucose production by suppressing Glucose-6-phosphatase and fructose-1,6-bisphosphatase enzyme activities [Chandramohan et al., 2008]. Since lipid abnormalities accompanying with premature atherosclerosis are the major causes of cardiovascular diseases in diabetic patients, ideal treatment for diabetes, in addition to glycemic control, should have a favorable effect on lipid profile. Cardiovascular diseases are listed as the cause of death in 65 % people suffering from diabetes [Kesari et al., 2007]. The liver is regarded as the central metabolic organ in the body, with an important role in glucose and lipid homeostasis [Eidi and Eidi, 2009]. The present study thus showed that the methanolic extract of *Pimenta dioica* leaf extract has potent antidiabetic properties in STZ induced diabetic rats which could be supplemented to such animals in different proportions depending upon the body weight.

## 5. REFERENCES

- Alluri V, Krishnaraj U (2006). Biological Screening of Medicinal Plants Collected from Eastern Ghats of India. *International Journal of Applied Science and Engineering* 4: 115-125
- Almdal TP (1988) Increased capacity of urea synthesis in streptozotocin diabetes in rats. *Diabetologia*. Nov;29(11):812-6.
- Al-yassin D, Ibrahim K (1981) A minor haemoglobin fraction and the level of fasting blood glucose. *J Fac Med Baghdad* 23:373–380.
- Bandaranayake, WM (2002) Bioactivities, bioactive compounds and chemical constituents of mangrove plants. *Wetlands Ecology and Management* 10 : pp. 421–452
- Beuchat, LR (1994). Antimicrobial properties of spices and their essential oils. In Dillon, Y.M., Board, R.G. (Eds.), *Natural Antimicrobial Systems and Food Preservation*. CAB International. Oxon PP. 167-179.
- Campos MG, Oropeza M, Torres-Sosa C, énez-Estrada M, Reyes-Chilpa R (2009) Sesquiterpenoids from antidiabetic *Psacalum decompositum* block ATP sensitive potassium channels. *J Ethnopharmacol* 123: 489-493
- Caraway WT (1955) Determination of uric acid in serum by carbonate. 25: 840-845.
- Chandramohan G, Ignaci muthu S, Pugalendi KV (2008) A novel compound from *Casearia esculenta* (Roxb.) root and its effect on carbohydrate metabolism in streptozotocin-diabetic rats. *Eur J Pharmacol* 590 (3): 437-443
- Chatterjee MN, Shinde R (2002) *Text Book of Medical Biochemistry*. New Delhi: Jaypee Brothers Medical Publishers.
- Chattopadhyay RR, Bandyopadhyay M (2005). Possible mechanism of hepatoprotective activity of *Azadirachta indica* leaf extract against paracetamol-induced hepatic damage in rats . Part III *Ind J.pharm.*37(3): 184-185.
- Craig ET (1997) Approaches and rationale for the design of synthetic antioxidants as therapeutic agents. In *Hand book of synthetic antioxidants*. ed. Lester pecker and Enrique cadenas. Marcel Decker pp, 1-52.
- Cutler HG (1995). Natural product flavor compounds as potential antimicrobials, insecticides and medicinal. *Agro- Food –Industry Hi-Tech*. 6, 19-23.
- Drabkin DL, Austin JM (1932). Spectrophotometer constants for common hemoglobin derivatives in human, dog and rabbit blood. *J. Biol.Chem* 98:719-733
- Eidi A, Eidi M (2009) Antidiabetic effects of sage (*Salvia officinalis* L.) leaves in normal and streptozotocin-induced diabetic rats. *Diabetes Met Syn Clin Res Rev* 3: 40-44.
- Fawcett JK, scott JE (1960) A rapid and precise method for the determination of urea. *J.Clin.path* 3:156-159.
- Gengaihi E, Souad, Nabaweya I, Sahar R, Regal E, Naglaa S. Sweet ent-kaurene (2011) diterpene glycosides of *Stevia rebaudiana* Leaves bertonii and biological evaluation. *J Amer Sci* 7: 775-782
- Gupta R, Bajpai KG, Johri S, Saxena AM (2007) An overview of indian novel traditional medicinal plants with anti-diabetic potentials. *Afr J Tradit Complement Altren med* 5: 1-17.
- Irshad M, Chaudhuri PS (2002) Oxidant, antioxidant system: Role and Significance in human body. *Ind J expt boil* 40(11): 1233-1239
- Jaffe M (1886) Concerning the precipitate produced in normal urine by picric acid and a new reaction of creatinine. *Z physiol.Chem.*10:139-400.
- Jarald EE, Joshi SB, Jain DC (2008) Antidiabetic activity of flower buds of *Michelia champaca* Linn. *Indian J Pharmacol* 40(6): 256-260
- Kaleem M, Sheema, Sarmad H, Bano Bn (2005) Protective effects of piper nigrum and vinca rosea in alloxan induced diabetic rats. *Indian J Physiol Pharmacol* 49(1):65–71 .
- Kesari AN, Kesari S, Singh SK, Gupta RK, Watal G (2007) Studies on the glycemic and lipidemic effect of *Murraya koenigii* in experimental animals. *J Ethnopharmacol* 112 (2): 305-311.
- Khan A, Zaman G, Anderson RA (2009) Bay leaves improve glucose and lipid profile of people with Type 2 diabetes. *J. Clin.Biochem. Nutr* 44: 52–56.
- Knentz AJ, Natras M (1991) Diabetic ketoacidosis, nonketotic hyperosmolar coma and lactic acidosis; *Handbook of diabetes*, eds. By Pickup JC, Williams G. Blackwell Science (2): 479–494.
- Koenig RJ, Peterson CM, Jones RL, Saudek C, Lehrman M, Cerami A (1976) Correlation of glucose regulation and haemoglobin A1c in diabetes mellitus. *New Engl J Med* 295(8):417–420.
- Kumar PJ, Clark M (2002) *Textbook of Clinical Medicine*. Saunders, London. 1099–1121.
- Marles RJ, Farnsworth N (1995) Antidiabetic plants and their act constituents. *Phytomedicine* 2(2):137–189.
- Nakatani, N (1994). Antioxidative and antimicrobial constituents of herbs and spices. In: charablambous, G. (Ed.), *Spices, Herbs and Edible Fungi*. Elsevier Science, New York PP. 251-271.
- Parab RS, Mengi SA (2002) Hypolipidemic activity of *Acorus calamus* L. in rats. *Fitoterapia* 73(6): 451-455.
- Pul ok K, Mukherjee (2002) *Quality control of Herbal Drugs* 1st edition. Business Horionel 2-21.

- Rang HP, Dale MM, Ritters JM (1991) The endocrine pancreas and the control of blood glucose; in Pharmacology; eds. By Simmons B, Beasley S. U.K. Longman group Ltd., 1991; 403–410.
- Santhakumari P, Prakasam A, Pugalendi KV (2006) Antihyperglycemic Activity of Piper betle Leaf on Streptozotocin-Induced Diabetic Rats. *J Med Food* 9(1):108–112.
- Sasaki T, Mastu S, Sonae A (1972) Effect of acetic acid concentration on the color reaction in the O-toluidine boric acid method for blood glucose estimation *Rinsho Kagaku* 1:346-353
- Saudek CD, Derr, R L, Kalyani RR (2006) Assessing glycemia in diabetes using self-monitoring blood glucose and hemoglobin HbA<sub>1c</sub>. *JAMA*.295:1688-1697.
- Shirwaikar A (2006) In vitro antioxidant studies on the benzyl tetra isoquinoline alkaloid berberine. *Biol Pharm Bull*. Sep;29(9):1906-10
- Shirwaikar A, Rajendran K, Barik R (2006) Effect of aqueous bark extract of *Garuga pinnata* Roxb. In streptozotocin–nicotinamide induced type II diabetes mellitus. *J. Ethnopharmacology* 107: 285–90.
- Shirwaikar A, Rajendran K, Dinesh Kumar C, Bodla R (2004) Antidiabetic activity of aqueous leaf extract of *Annona squamosa* in streptozotocin nicotinamide type 2 diabetic rats. *J. Ethnopharmacology* 91:171–5.
- Sudhakar Nayak S, Pattabiraman TN (1981) A new colorimetric method for the estimation of glycosylated hemoglobin. *Clin Chin Acta* 109:267–274.
- Tietz NW (1987). *Fundamentals of clinical Chemistry*. WB Saunders Company Philadelphia PP.638
- wanston-Flat SK, Day C, Bailey CJ, Flatt PR (1990) Traditional plant treatment for diabetes: studies in normal and streptozotocin diabetic mice. *Diabetologia* 33:462–464.

\*\*\*\*\*